



The 65th ASH Annual Meeting Abstracts

POSTER ABSTRACTS

722.ALLOGENEIC TRANSPLANTATION: ACUTE AND CHRONIC GVHD, IMMUNE RECONSTITUTION

Improved T- and B-Cell Neogenesis in Children with Acute Leukemia Given an Alpha-Beta T-Cell Depleted Haplo-HSCT Combined with the Infusion of Donor T-Cells Genetically Modified with Inducible Caspase 9 Suicide Gene (Rivo-cel)

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Background

$\alpha\beta$ T- and B-cell depleted HLA-haploidentical HSCT ($\alpha\beta$ haplo-HSCT) is a suitable transplant option promptly available and virtually applicable to any children in need of an allograft. However, the suboptimal recovery of adaptive immunity still represents a limitation of the approach. In order to better deconvolute the reconstitution of adaptive immunity, in this study we measured T-cell receptor and kappa chain-deleting recombination excision circles (TREC and KREC), which represent reliable surrogates of T and B-cell neogenesis, respectively. We also analyzed a cohort of patients who received after the allograft infusion of a titrated number of donor T-cells genetically modified with inducible caspase 9 (iC9) suicide gene (Rivo-cel) as part of a prospective clinical trial (NCT02065869) with the goal of accelerating the recovery of thymus-independent adaptive immunity.

Patients and study design

We retrospectively analyzed 134 patients (≤ 25 years) with acute lymphoblastic (ALL) or myeloid leukemia (AML) who received a $\alpha\beta$ haplo-HSCT between 2014 and 2021; 65 of them received post-transplant infusion of Rivo-cel, at the dose of 1×10^6 cells/kg, after a median time of 26 days (range: 11-87) post-transplant. No patient received post-transplantation pharmacological GVHD prophylaxis (Locatelli et al. Blood 2017). Details on patient and donor characteristics, as well as on graft composition are reported in Table 1.

T-cell neogenesis was investigated by quantifying signal joint- (sj-) and beta (b-) TREC while B-cell neogenesis was analyzed by measuring coding-joint (cj-) and sj- KREC. We performed real-time quantitative PCR, as previously described (Arruda et al. 2018), on genomic DNA extracted from PBMC collected at 6 different time points (before and 1, 3, 6, 12 and 18 months after the allograft). At the same time-points peripheral blood of patients was also evaluated by flow-cytometry to phenotypically characterize the pool of leukocytes.

Results

With a median follow-up of 6 years (range 1-9) the overall (OS) and disease-free survival (DFS) probabilities of the entire cohort were 88,1% and 82%, respectively. The cumulative incidence (CI) of relapse was 15%. CI of grade II-IV acute Graft-versus-Host Disease (GvHD) was 19%, while that of chronic GvHD (cGvHD) was 7%. In the Rivo-cel cohort, sjTREC and bTREC were consistently detectable as early as one month post-HSCT, while started to recover by 3 months after $\alpha\beta$ haplo-HSCT alone, and were significantly higher at +3 ($p=0,0006$ and $p=0,02$, respectively) and +6 ($p=0,0003$ and $p=0,001$) months post-HSCT as compared with $\alpha\beta$ haplo-HSCT. sjTREC and bTREC reached pre-transplant values at 6 and 12 months in Rivo-cel and $\alpha\beta$ haplo-HSCT cohort, respectively. B-cell neogenesis, assessed by sj and cjKRECs, showed a superimposable recovery kinetics. A higher number of sjKRECs was observed in Rivo-cel cohort at +1, +3 and +6 months after transplantation as compared with $\alpha\beta$ haplo-HSCT ($p=0,007$, $p=0,02$ and $p=0,04$ for sjKRECs; $p=0,005$, $p=0,092$ and $p=0,035$ for cjKRECs, respectively). Furthermore, in the Rivo-cel cohort, patients with detectable sj and bTREC values at +3 months after transplant showed a significantly higher overall survival ($p=0,01$ and $p=0,02$, respectively). Also in this cohort, aGvHD was confirmed to play a detrimental effect on T-cell neogenesis, at 3 and 6 months after transplant. The same negative effect was documented for chronic GvHD at +6, +12 and +18 months, also affecting B-cell neogenesis at +6 months after transplant. Type of leukemia did not affect T and B-cell neogenesis.

Conclusions

In patients given $\alpha\beta$ haplo-HSCT, T-cell and B-cell neogenesis were shown to be significantly accelerated by the post-transplant infusion of a titrated number of donor T-cells genetically modified with the iC9 suicide gene. Further studies are needed to elucidate the biologic mechanisms responsible for this favorable effect on recovery of adaptive immunity. Our data confirm the detrimental effect of both acute and chronic GvHD in recovery of thymic function and B-cell neogenesis. In this perspective, the possibility to successfully control/abrogate GvHD through the apoptosis of Rivo-cel triggered by the infusion of the dimerizing agent (AP1903) offers also the advantage of, at least partly, preventing the detrimental effect played by this complication on immune recovery.

Disclosures Algeri: Vertex Pharmaceuticals: Consultancy, Membership on an entity's Board of Directors or advisory committees. **Merli:** JAZZ: Consultancy, Honoraria; MEDAC: Speakers Bureau; SOBI: Consultancy; Amgen: Speakers Bureau. **Locatelli:** Bellicum, Amgen, Neovii, Novartis. Sanofi, SOBI, Vertex: Membership on an entity's Board of Directors or advisory committees, Speakers Bureau; Miltenyi, Jazz Pharm, Medac, Sobi, Gilead, BluebirdBio: Speakers Bureau; Sanofi, Vertex: Membership on an entity's Board of Directors or advisory committees.

Table 1

	Patients (n=134)	$\alpha\beta$ Haplo-HSCT + Rivo-cel (n=65)	$\alpha\beta$ Haplo-HSCT (n=69)	
Sex				
Male	78	35	43	N.S.
Female	56	30	26	
Median (range) at diagnosis, y				
Median (range) at transplantation, y	6,8 (0-20,5)	5,8 (0-17,7)	7,8 (0,2-20,5)	N.S.
Diagnosis				<i>p=0,04</i>
LLA	94	40	54	
LMA	40	25	15	
Disease status at transplantation				
ALL				
CR1	25	10	15	
CR2	63	30	33	
≥ CR3	6	0	6	
AML				
CR1	31	21	10	
CR2	9	4	5	
Conditioning Regimen				
TBI-based	100	44	56	N.S.
Chemo-based	34	21	13	
Anti-T-lymphocyte globulin (from day -5 to -3)	134	65	69	
Rituximab (day -1)	134	65	69	
Donor characteristics				
Type of donor				
Mother/Father	63/62	32/27	31/35	N.S.
Brother/Sister	6/3	3/3	3/0	
Median age (range), y	43 (19-56)	37 (19-50)	43 (21-56)	<i>p=0,001</i>
Cell dose infused, median (range)				
CD 34+ cells x 10 ⁶ /Kg	16,6 (3,48-38,79)	18 (3,48-38,79)	15,49 (6-33,84)	<i>p=0,02</i>
CD $\alpha\beta$ + cells x 10 ⁶ /Kg	0,036 (0,003-0,099)	0,033 (0,004-0,097)	0,039 (0,003-0,099)	
CD $\gamma\delta$ + cells x 10 ⁶ /Kg	7,4 (0,86-56,7)	9,4 (0,86-38,9)	6,9 (1,35-56,7)	

Figure 1

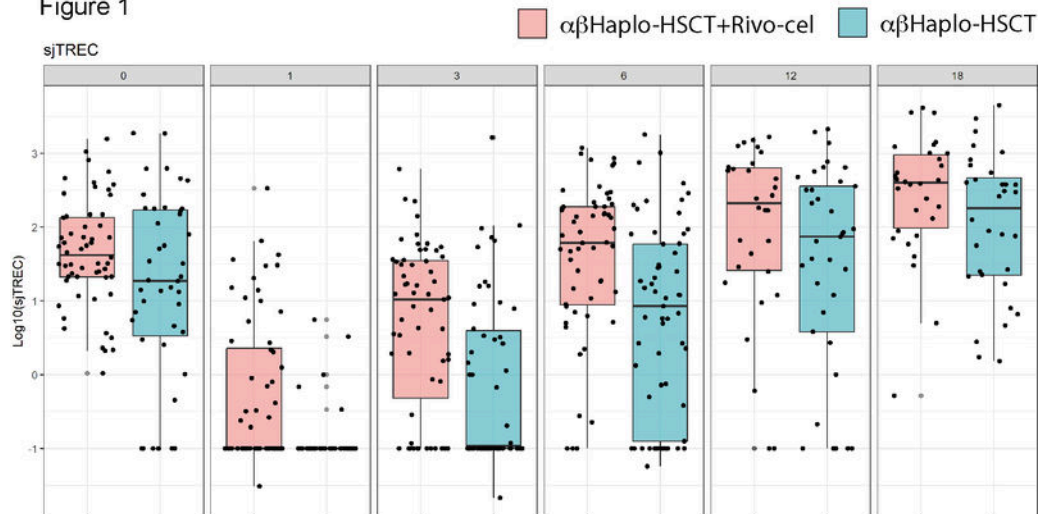


Figure 1

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